



skyla™ Basic Biochemistry Panel

HB1

IVD

PN : 800-110

For In Vitro Diagnostic Use and For Professional Use Only

Rev : E

1. Intended Use

The skyla Basic Biochemistry Panel used with skyla Clinical Chemistry Analyzer, is intended to be used for the quantitative determination of Albumin (ALB), Alanine Aminotransferase (ALT/GPT), Blood Urea Nitrogen (BUN), Creatinine (CREA), Blood Glucose (GLU), Total Protein (TP), Uric Acid (UA) in human whole blood, plasma, or serum. The calculated values of Albumin/Globulin Ratio (A/G Ratio), Estimated Glomerular Filtration Rate (eGFR), and c Globulin (GLOB) can then be obtained.

2. Principles

The skyla Basic Biochemistry Panel contains a total of 7 types of dried reagents located in the respective detection wells of the reagent disc. The user only needs to inject the blood specimens into the sample port of the disc, and places the disc into the analyzer. The test will be done automatically within 15 minutes. Three additional calculated values are also obtained after the test. For the detail description of disc, please refer to “skyla Clinical Chemistry Analyzer Operator’s Manual”.

Clinical Significance:

Albumin (ALB)

ALB is the major protein component of normal human serum, accounting for more than 50% of the total protein. It plays an important role in the regulation of the osmotic blood pressure. Abnormal ALB values may be caused by dehydration, malnutrition, nephrotic syndrome or liver dysfunction.

Alanine Aminotransferase (ALT/GPT)

ALT is one of the indicators of liver function. Acute and chronic hepatitis, drug induced liver injury, fatty liver, cirrhosis, myocardial infarction, myocarditis and biliary diseases can lead to elevated ALT activity.

Blood Urea Nitrogen (BUN)

BUN is one of the important markers for diagnosis and prognosis tracking of kidney diseases and metabolic disorders. Other common possible causes of elevated BUN include dehydration and heart failure.

Creatinine (CREA)

CREA is the degradation product of creatine in human muscles. It is a commonly used marker to examine renal functions. Elevated CREA in the blood may be caused by severe muscle disease, nephritis, hyperthyroidism and malnutrition.

Glucose (GLU)

GLU can be used for the diagnosis of diabetes and diseases related to the carbohydrate metabolism. Diabetes, chronic pancreatitis and certain endocrine diseases may lead to hyperglycemia. Abnormal glucose metabolism, islet cell tumors, pancreatic tumors and severe liver diseases may lead to hypoglycemia.

Total Protein (TP)

TP is an indicator for the liver function and kidney diseases. Elevated TP could be caused by dehydration or increased immunoglobulin levels. And TP reduction may occur in the disorders include malnutrition, nephrotic syndrome, various liver diseases and malignant tumors.

Uric Acid (UA)

UA can be used for diagnosis and prognosis tracking of kidney related diseases and diseases caused by metabolic disorders. Kidney diseases, lactic acidosis, dehydration, preeclampsia, or diabetic ketoacidosis can lead to increased UA concentration.

Albumin/Globulin Ratio (A/G Ratio)

The A/G Ratio is the ALB and GLOB ratio. It is used to assess liver function and is an important indicator for the diagnosis of viral hepatitis and cirrhosis.

Estimated Glomerular Filtration Rate (eGFR)

eGFR is the kidney filtrate per minute, which is calculated from CREA. It is used to assess renal function.

Globulin (GLOB)

It is calculated from TP and ALB. It is used to assess liver function.

Method:

ALB

ALB is determined through the endpoint color reaction method. When binding to Bromocresol Green (BCG), it forms a yellow-green complex. The absorbance at a wavelength of 600nm can be measured. The amount of ALB in the sample is proportional to the bound ALB.

ALT

ALT activity is enzymatically determined. ALT catalyses the reaction of Alanine with α -Ketoglutarate, converting them into Glutamate and Pyruvate. In the presence of NADH, Lactate Dehydrogenase converts Pyruvate into Lactate. In the course of the reaction NADH is oxidized to NAD. The decrease of NADH absorbance is measured at a wavelength of 340 nm and is proportional to ALT activity.

BUN

BUN is enzymatically determined. Urea undergoes a Urease catalyzed hydrolysis, thus producing Ammonia and Carbon Dioxide. In a Glutamate Dehydrogenase (GLDH) catalyzed reaction Ammonia reacts with 2-Oxoglutarate yielding L-Glutamate. In the process of this reaction, NADH is oxidized to NAD which in turn undergoes a color reaction. The rate of change of absorbance at wavelength of 340 nm is measured and proportional to the BUN concentration.

CREA

CREA is determined through the endpoint enzymatic reaction approach. Creatinine Amidohydrolase hydrolyzes CREA to Creatine. Then Creatine is converted into Sarcosine through catalysis of Creatine Amidohydrolase. Furthermore, Sarcosine Oxidase oxidizes Sarcosine, yielding Glycine, Formalehyde and Peroxide (H_2O_2) in the process. The enzyme Peroxidase processes Hydrogen Peroxide, 2,4,6-3 Hydroxy-Benzoic acid (TBHBA) and 4-Amine Triazolam Alternate Pyrazol (4-AAP), forming a Quinoneimine dye as a product. The dye formation is measured at wavelength of 546 nm and is proportional to the amount of CREA in the sample.

GLU

GLU is determined through the endpoint enzymatic reaction approach. The Sucrose is catalyzed by Hexokinase to D-Glucose-6-Phosphate (G-6-P). In the presence of NAD, G-6-PD converts G-6-P into 6- Phosphogluconate and NADH. The absorbance at the wavelength of 340 nm can be measured in the presence of NADH. The absorbance is proportional to the GLU concentration.

TP

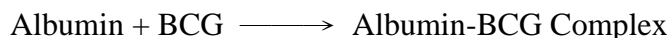
TP is determined by the Biuret method. The peptide bonds of the protein react with Copper ions in an alkaline environment and form a purple compound. The color development is proportional to the original TP concentration and is measured at wavelength of 546 nm.

UA

UA is determined enzymatically by an endpoint reaction. In this method, Uric Acid is converted into Allantoin and Peroxide. The Peroxidase catalyzed reaction of Peroxide with 4-Aminoantipyrine (4-AAP) and 3,5-Dichloro+2-Hydroxybenzene-Sulfonate (DCHBS) results in the formation of a Quinoneimine dye. The dye formation is proportional to the UA concentration and is measured at wavelength of 510 nm.

Reaction pathway :

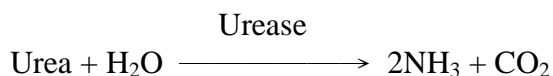
ALB



ALT

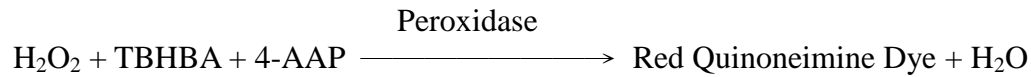
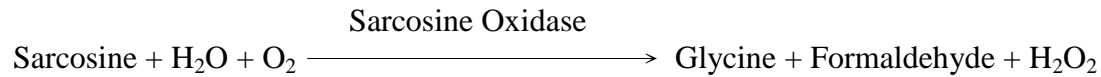
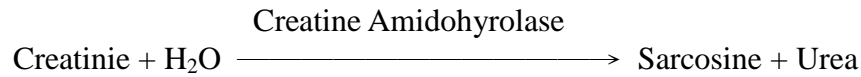
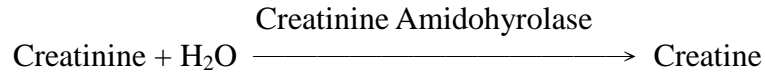


BUN

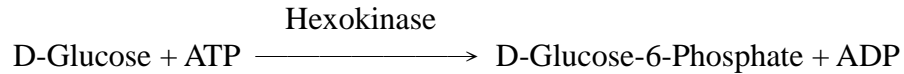




CREA



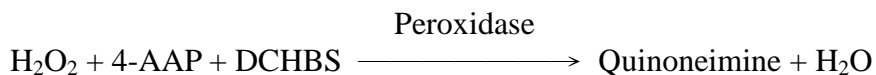
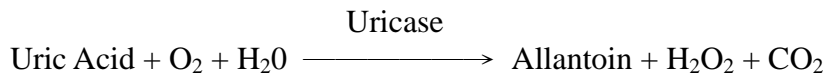
GLU



TP



UA



3. Reagents

Included:

Each panel contains dried reagent beads, dried internal QC beads and the diluent.

Reagent Composition:

| Composition | Quantity/Panel |
|-------------------------|----------------|
| 4-AAP | 0.04 mg |
| ATP | 0.04 mg |
| Bromocresol Green | 5.4 µg |
| Copper Sulfate | 0.1 mg |
| Creatinase | 2.8 U |
| Creatininase | 5.6 U |
| DCHBS | 0.1 mg |
| G6PDH | 0.2 U |
| Glutamate Dehydrogenase | 0.05 U |
| Hexokinase | 0.1 U |
| Lactate Dehydrogenase | 0.3 U |
| L-Alanine | 0.3 mg |
| NAD | 0.1 mg |

| Composition | Quantity/Panel |
|-----------------------------|----------------|
| NADH | 0.06 mg |
| Peroxidase | 0.7 U |
| Sarcosine Oxidase | 0.4 U |
| TBHBA | 0.2 mg |
| Urease | 0.03 U |
| Uricase | 0.3 U |
| α -Ketoglutaric Acid | 0.25 mg |

Reagent Storage:

- The reagent disc should be stored at 2~8°C.
- The expiry date of the reagent is printed on the outside of the sealed pouch of reagent disc. Do not use if the reagents have expired.

4. Specimen Collection and Preparation

Specimen Collection:

- Specimens suitable for skyla Basic Biochemistry Panel include lithium heparinized whole blood, lithium heparinized plasma, serum and quality control solutions. The sample requirement is 200 μ L. (\pm 10 μ L tolerance are allowable)
- Collection, preservation and handling of specimens in accordance with local legal requirements or the standard operating procedures of your organization.

Note: Do not use specimens containing other coagulants. That would cause in incorrect test results.

Specimen Preparation:

- Before applying a sample to the reagent disc, gently rotate the sample tube up and down several times, to confirm the sample is homogeneous (evenly mixed). If the sample is whole blood, do not shake the sample container vigorously to avoid occurrence of hemolysis.

Note: 1. Perform testing within 10 minutes after applying the sample to the reagent disc.

2. The use of whole blood specimens with hematocrits (Hct) higher than 60% may affect the test results.

For further information in specimen collection and preparation, please refer to “skyla Clinical Chemistry Analyzer Operator’s Manual”.

5. Test Procedures

Material Preparation:

1 piece of the reagent disc of skyla Basic Biochemistry Panel

Required materials not included in the panel:

The skyla Clinical Chemistry Analyzer

Sample collection container

Micropipette / Tips

Control reagents available on the market.

Test Conditions:

Test should be carry out in an environment with temperatures of 10°C~32°C. Each test will take about 15 minutes. During the test, chamber in the analyzer keeps the temperature at 37°C for stable assay.

Test Steps:

1. Open the aluminum pouch and remove the reagent disc.
2. Remove the diluent container sealing.
3. Using a micropipette to inject 200µL of the sample into the reagent disc through the sample port.
4. Place the reagent disc to the analyzer drawer.
5. Press the “start” button on the screen to initiate testing.

For details on the operating steps and instrument setting, refer to “the skyla™ Clinical Chemistry Analyzer Operator’s Manual.”

- Note:
1. To operate the reagent disc or instrument, please wear lab gloves and other protective gear to avoid contamination by specimen.
 2. The used reagent disc, tips should be discarded as biomedical waste.
 3. Testing should be performed within 20 minutes after the pouch is opened.
 4. Do not place the reagent disc at the environment more than 25°C and longer than 48 hours prior to use.
 5. If the reagent disc or its package is damaged or is over the expiry date, do not use it.

6. Calibration

The barcode on every manufactured reagent disc contains all information required for calibration of the test items. The analyzer will automatically read the barcode information during testing.

7. Quality Control

External quality control materials can be used for the accuracy monitor of skyla system. The recommended frequency of QC testing is as follows. (External quality control materials are not provided by LITE-ON)

- At least every 30 days.
- Before a new batch of reagents is used for testing.
- When the analyzer is moved or the operating environment significantly changes.

8. Reference interval

The table below shows the reference interval for each test item. These ranges are provided as a reference only. It is recommended that every laboratory or test site should establish its own reference interval from its particular patient population.

| Test Item | Reference Interval | Reference Interval (SI Unit) | |
|-----------|--------------------|------------------------------|------------------|
| ALB | 3.5 – 5.3 g/dL | 35 – 53 g/L | |
| ALT | < 40 U/L | < 40 U/L | |
| BUN | 9 – 23 mg/dL | 3.2 – 8.2 mmol urea/L | |
| CREA | Male | 0.7 – 1.3 mg/dL | 62 – 115 µmol/L |
| | Female | 0.6 – 1.1 mg/dL | 53 – 97.2 µmol/L |
| GLU | 70 – 110 mg/dL | 3.9 – 6.1 mmol/L | |

| Test Item | Reference Interval | Reference Interval (SI Unit) |
|-----------|--------------------|------------------------------|
| TP | 6.0 – 8.3 g/dL | 60 – 83 g/L |
| UA | Male | 238 – 446 μ mol/L |
| | Female | 178 – 357 μ mol/L |

9. Limitation

Interference studies:

1. Effect of endogenous substances

Physiological interferents in blood include hemolysis, icterus, and lipemia. For every test item, 2 Levels human serum pool supplemented with known concentrations of the endogenous substances were used for the testing. Significant interference is defined as a >10% shift in the test result.

| Test Item | substance concentration with interferences of less than 10% | | | |
|-----------|---|---------------------------------------|-------------------------------------|-------------------------|
| | Hemolysis [Hemoglobin] | Icterus [Bilirubin (unconjugated)] | Icterus [Bilirubin (conjugated)] | Lipemia [Intralipid] |
| ALB | 147.6 mg/dL | 62.5 mg/dL | 57.5 mg/dL | 0.11% |
| ALT | 290 mg/dL | 43.5 mg/dL | 22.3 mg/dL | 0.02% |
| BUN | 522.5 mg/dL | 50.5 mg/dL | 34.8 mg/dL | 0.1% |
| CREA | 170 mg/dL | 5.2 mg/dL | 1.2 mg/dL | 0.12% |
| GLU | 600 mg/dL | 62.5 mg/dL | 55.5 mg/dL | 0.017% |
| TP | 157.2 mg/dL | 62.5 mg/dL | 57.5 mg/dL | 0.07% |
| UA | 253.1 mg/dL | 9.8 mg/dL | 6.26 mg/dL | 0.03% |

2. Effect of exogenous substances

Ten exogenous substances were selected as potential interferents for the study. For every test item, human serum pool supplemented with a known concentration of the substances was used for the testing. Significant interference is defined as a >10% shift in the test result.

| Substance | Test Concentration | Affected Test Item | Effect |
|----------------------|--------------------|-----------------------------|------------|
| Acetaminophen | 20 mg/dL | No significant interference | |
| Acetylsalicylic acid | 65 mg/dL | No significant interference | |
| Ampicillin | 5 mg/dL | No significant interference | |
| Ascorbic acid | 6 mg/dL | UA | 15.7% Dec. |
| Caffeine | 6 mg/dL | No significant interference | |
| Cephalothin | 30 mg/dL | No significant interference | |
| Cimetidine | 2 mg/dL | No significant interference | |
| Ibuprofen | 50 mg/dL | CREA | 12.7% Inc. |
| Salicylic acid | 60 mg/dL | CREA | 12.7% Inc. |
| Theophylline | 4 mg/dL | No significant interference | |

10. Performance Characteristics

Dynamic range:

The dynamic range was determined by linearity study, as follows:

| Test Item | Dynamic Range | Dynamic Range (SI Unit) |
|-----------|----------------|-------------------------|
| ALB | 1.0 – 6.0 g/dL | 10 – 60 g/L |
| ALT | 20 – 1100 U/L | 20 – 1100 U/L |
| BUN | 2 – 120 mg/dL | 0.7 – 42.8 mmol urea/L |
| CREA | 0.6 – 20 mg/dL | 53 – 1768 µmol/L |
| GLU | 30 – 600 mg/dL | 1.7 – 33.3 mmol/L |
| TP | 1.5 – 10 g/dL | 15 – 100 g/L |
| UA | 1 – 20 mg/dL | 59 – 1190 µmol/L |

Analytical Sensitivity:

The sensitivity (limits of quantitation) was determined according to the lowest concentration of the dynamic range which had an acceptable CV (CV<20%). The sensitivity of each test item is shown in the table below.

| Test Item | Limit of Detection | Test Item | Limit of Detection |
|-----------|--------------------|-----------|--------------------|
| ALB | 1.0 g/dL | GLU | 30 mg/dL |
| ALT | 20 U/L | TP | 1.5 g/dL |
| BUN | 2 mg/dL | UA | 1 mg/dL |
| CREA | 0.6 mg/dL | | |

Precision:

Precision studies adopt serum pool of high and low concentrations as test samples. Tests are performed twice a day for a total of 20 days. Results for repeatability and reproducibility of each test item are shown in the table below.

| Level 1 | | | | | |
|-----------|-------------|------------|------------|-------|------------|
| Test Item | Mean | Within-Run | | Total | |
| | | SD | %CV | SD | %CV |
| ALB | 4.89 g/dL | 0.09 | 1.8 | 0.09 | 1.8 |
| ALT | 54.0 U/L | 1.6 | 3.0 | 1.7 | 3.1 |
| BUN | 14.46 mg/dL | 0.52 | 3.6 | 0.55 | 3.8 |
| CREA | 3.0 mg/dL | 0.09 | 2.9 | 0.11 | 3.6 |
| GLU | 84.7 mg/dL | 1.4 | 1.6 | 1.4 | 1.7 |
| TP | 6.65 g/dL | 0.07 | 1.0 | 0.07 | 1.0 |
| UA | 3.25 mg/dL | 0.14 | 4.4 | 0.14 | 4.4 |

| Level 2 | | | | | |
|-----------|-------------|------------|------------|-------|------------|
| Test Item | Mean | Within-Run | | Total | |
| | | SD | %CV | SD | %CV |
| ALB | 2.56 g/dL | 0.05 | 2.1 | 0.06 | 2.2 |
| ALT | 194.6 U/L | 6.1 | 3.1 | 6.3 | 3.2 |
| BUN | 23.29 mg/dL | 0.73 | 3.1 | 0.79 | 3.4 |
| CREA | 7.5 mg/dL | 0.32 | 4.3 | 0.32 | 4.3 |
| GLU | 274.7 mg/dL | 2.4 | 0.9 | 3.2 | 1.1 |
| TP | 4.16 g/dL | 0.06 | 1.4 | 0.06 | 1.5 |
| UA | 6.52 mg/dL | 0.21 | 3.2 | 0.22 | 3.4 |

Method Comparison:

The automatic clinical chemistry analyzer in clinical laboratory was used as comparative method in the study. The tests are performed by using the same clinical serum sample for two methods. Correlation between two methods can be determined through statistical analysis.

| Test Item | Correlation | | | SEE | N | Sample range |
|-----------|-----------------|-------|-----------|-------|----|--------------------|
| | Coefficient (R) | Slope | Intercept | | | |
| ALB | 0.9850 | 1.008 | -0.015 | 0.148 | 52 | 1.63 – 5.34 g/dL |
| ALT | 0.9995 | 1.019 | 0.9 | 5.1 | 44 | 4 – 807 U/L |
| BUN | 0.9976 | 0.994 | 0.248 | 1.325 | 53 | 3.9 – 106.3 mg/dL |
| CREA | 0.9979 | 0.895 | 0.142 | 0.231 | 40 | 0.21 – 18.11 mg/dL |
| GLU | 0.9986 | 1.004 | 0.2 | 6.3 | 56 | 32 – 640 mg/dL |
| TP | 0.9911 | 0.999 | -0.008 | 0.202 | 52 | 2.36 – 9.34 g/dL |
| UA | 0.9967 | 1.012 | -0.048 | 0.254 | 63 | 2.02 – 18.57 mg/dL |

Matrix Comparison:












The Correlation between WB, plasma and serum was determined. The clinical sample was used in the study.

| Test Item | N | Matrix type | Correlation | | |
|-----------|---|------------------|-----------------|-------|-----------|
| | | | Coefficient (R) | Slope | Intercept |
| ALB | 5 | Serum vs. Plasma | 0.9949 | 1.000 | -0.04 |
| | | Plasma vs. WB | 0.9999 | 1.005 | -0.10 |
| | | WB vs. Serum | 0.9961 | 0.996 | 0.14 |

| Test Item | N | Matrix type | Correlation Coefficient (R) | Slope | Intercept |
|-----------|----|------------------|-----------------------------|-------|-----------|
| ALT | 11 | Serum vs. Plasma | 0.9991 | 0.984 | -0.261 |
| | | Plasma vs. WB | 0.9980 | 0.996 | -1.003 |
| | | WB vs. Serum | 0.9989 | 1.020 | 1.288 |
| BUN | 11 | Serum vs. Plasma | 0.9971 | 1.011 | -0.129 |
| | | Plasma vs. WB | 0.9996 | 1.006 | -0.344 |
| | | WB vs. Serum | 0.9967 | 0.983 | 0.466 |
| CREA | 11 | Serum vs. Plasma | 0.9991 | 0.999 | 0.035 |
| | | Plasma vs. WB | 0.9995 | 1.006 | -0.005 |
| | | WB vs. Serum | 0.9999 | 0.994 | -0.030 |
| GLU | 15 | Serum vs. Plasma | 0.9831 | 1.002 | 2.040 |
| | | Plasma vs. WB | 0.9851 | 1.060 | -3.935 |
| | | WB vs. Serum | 0.9911 | 0.941 | 1.619 |
| TP | 15 | Serum vs. Plasma | 0.9926 | 0.967 | 0.325 |
| | | Plasma vs. WB | 0.9965 | 1.038 | -0.188 |
| | | WB vs. Serum | 0.9960 | 0.996 | -0.148 |
| UA | 13 | Serum vs. Plasma | 0.9958 | 0.988 | 0.239 |
| | | Plasma vs. WB | 0.9979 | 1.050 | -0.244 |
| | | WB vs. Serum | 0.9971 | 0.964 | -0.006 |

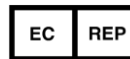
11. Reference

1. Clinical and Laboratory Standards Institute. Interference Testing in Clinical Chemistry; Approved Guideline - Second Edition. CLSI document EP07-A2. Robert J. McEnroe: 2005.
2. Clinical and Laboratory Standards Institute. Evaluation of the Linearity of Quantitative Measurement Procedures: A Statistical Approach; Approved Guideline. NCCLS document EP06-A. Dan Tholen: 2003.
3. Clinical and Laboratory Standards Institute. Protocols for Determination of Limits of Detection and Limits of Quantitation; Approved Guideline. NCCLS document EP17-A. Daniel W. Tholen: 2004.
4. Clinical and Laboratory Standards Institute. Evaluation of Precision Performance of Quantitative Measurement Methods; Approved Guideline - Second Edition. NCCLS document EP05-A2. Jan S. Krouwer: 2004.
5. Clinical and Laboratory Standards Institute. Method Comparison and Bias Estimation Using Patient Samples; Approved Guideline - Second Edition. NCCLS document EP09-A2. Jan S. Krouwer: 2002.

| Symbol Index | | | |
|---|------------------------------------|---|---|
|  | Catalogue number |  | Consult instruction for use |
|  | Batch code |  | Use by |
|  | Manufacturer |  | Authorized representative in the European Community |
|  | In Vitro diagnostic medical device |  | CE mark |
|  | Temperature limitation |  | Caution |
|  | Do not reuse | | |



LITE-ON Technology Corporation H.S.P.B.
No. 8, Dusing Road, Hsinchu Science Park
Hsinchu, Taiwan



MT Promedt Consulting GmbH
Altenhofstr. 80
D-66386 St. Ingbert
Germany

Customer service/Technical support : +886-3-611-8511
Email : support@skyla.com
Website : www.skyla.com

Issue date: 1 JUL 2013
Revise date: 18 APR 2016
PN: 7B25000034HD
LITE-ON Technology Corp.